Fusarium Research Center Culture Collection
Department of Plant Pathology
118 Buckhout Laboratory
The Pennsylvania State University
University Park, Pennsylvania 16802
U.S.A.

The culture you have received has been grown on carnation leaf-water agar. After 7-10 days growth on this medium, several colonized carnation leaf pieces were removed and lyophilized in sterile skim milk. The method and the use of irradiated carnation leaves in this process are described in the reference given below.

Fisher, N. L., L. W. Burgess, T. A. Toussoun, and P. E. Nelson. 1982. Carnation leaves as a substrate and for preserving cultures of *Fusarium* species. Phytopathology 72:151-153.

To revive this culture place the entire pellet on a suitable medium, such as carnation leaf-water agar, straw-water agar or another medium low in carbohydrates. Potato-dextrose agar may also be used but since some *Fusarium* species mutate rapidly in culture, a low carbohydrate medium is preferable. The entire pellet should be used because the carnation leaf pieces may not be uniformly distributed throughout the pellet. Growth from the pellet usually begins within 24 to 48 hours after it is placed on the growth medium but some cultures grow more slowly and it is necessary to wait 6 or 7 days before growth is evident. As the dried skim milk begins to liquefy, it may be mistaken for colonies of bacteria. All cultures are examined after lyophilization and bacterial contamination during storage is unlikely.

If you have any trouble reviving or growing this culture, please contact Dr. David Geiser (814 865-9773/ dgeiser@psu.edu) or Jean Juba (814-863-0145/ jhj2@psu.edu). Information on the preparation and use of natural media low in carbohydrates is given in the references below.

Hansen, H. N., and W. C. Snyder. 1947. Gaseous sterilization of biological materials for use as culture media. Phytopathology 37:369-371.

Snyder, W. C., and H. N. Hansen. 1947. Advantages of natural media and environments in the culture of fungi. Phytopathology 37:420-421.