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Dear Kevin,

I'm depositing five <u>Neurospora crassa</u> strains (MB 5, 17, 19, 26 and 27) that are all RIP'd mutants of a fatty acid synthase (FAS) gene. The proposed designation for these mutants is <u>cel-2</u>. The particular RIP'd gene is the one showing homology to the fatty acid synthase (beta subunit) gene of <u>Saccharomyces cerevisiae</u> (FAS1) and other fungi.

To RIP the gene, I used a plasmid provided by Dorsey Stuart's lab. I assume it is the same one identified out of the Stock Center's lambda-ZAP library that is mentioned by R. Yamashita and D. Stuart in the Fungal Genetics Newsletter (#43, pp. 66-67). The plasmid contains a portion of the gene that is about 3 kb of the 3' end (the complete <u>S. cerevisiae</u> gene is about 6 kb; Kottig et al., Mol. Gen. Genet. 226: 310-314). I cotransformed this plasmid into wild type with pBARMTE1 and selected for Ignite resistance, then checked stable transformants for the presence of a second copy of the beta-FAS gene by Southern blotting.

A transformant having a second copy was crossed to wild type, and single ascospores from the cross were grown on VMN + 1% Tween 40. Tween 40 provides palmitate, the optimal supplement for the <u>cel</u> strain, which synthesizes little fatty acid <u>de novo</u>. The <u>cel</u> strain, if it has a defective FAS, should have a defect in the alpha subunit (based on its low level of 4-phosphopantetheine in FAS, which is bound to the acyl carrier protein region of this subunit). Beta-subunit mutants were expected to have a phenotype similar to <u>cel</u>. Each of these 5 lines has a Tween 40 or palmitate supplementation requirement, showing virtually no growth without the supplement. The requirement is thus tighter than for <u>cel</u>, which grows very slowly without supplementation.

The ability of these lines to synthesize fatty acids <u>de novo</u> was determined by feeding them 30 μ M labelled (deuterated) palmitate, which we detect by GC/MS. These lines synthesize less than 10% of their total